

Available online at www.sciencedirect.com



Tetrahedron

Tetrahedron 63 (2007) 10077-10082

Synthesis of a transition-state analog for the hydrolysis of the zearalenone lactone

Krishanthi P. Jayasundera, Samuel J. Brodie and Carol M. Taylor*

Institute of Fundamental Sciences, Massey University, Private Bag 11-222, Palmerston North, New Zealand

Received 22 January 2007; revised 28 June 2007; accepted 12 July 2007 Available online 19 July 2007

Abstract—The development of a catalytic antibody for the hydrolysis of the lactone functionality in zearalenone (1) is viewed as a potential solution to animal fertility problems associated with the estrogenic mycotoxin. A phosphonomacrolactone is proposed as a hapten for the generation of such antibodies. A suitably functionalized aryl phosphonic acid 4 was condensed with the racemic aliphatic fragment 5 via a Mitsunobu reaction. Macrocyclic formation was achieved via RCM to give advanced intermediate 23, a phosphonate analog of zearalenone, ready for deprotection and conjugation to a carrier protein. © 2007 Elsevier Ltd. All rights reserved.

1. Introduction

Zearalenone (1) is a mycotoxin, first isolated by Stob and coworkers in 1962,¹ which is produced by various species of *Fusarium*.² Zearalenone is estrogenic and affects a variety of animals. In New Zealand, sheep ingests zearalenone when grazing on fungi-infested pastures.³ Levels of the toxin peak in the warm, dry conditions of late summer, and early autumn and unfortunately coincide with the sheep mating season. This results in reduced fertility: more barren ewes, fewer twins, later lambs, and associated productivity losses.

This toxin represents a considerable problem to the global agricultural and horticultural industries. It is not surprising that a number of methods have been considered to counter its effects. Degradation of the toxin to harmless byproducts has been demonstrated by microorganisms.⁴ Androvax[®] is a steroid-based vaccine that stimulates ovulation and thereby counteracts the detrimental effects of toxins such as zearal-enone.⁵ A novel approach would be the use of catalytic antibodies to degrade the toxin in vivo—either by inoculation with an antibody preparation or by stimulating the animals to produce their own therapeutic antibodies.

We proposed that a transition-state analog for hydrolysis of the macrolactone would be a suitable hapten for the production of such antibodies (Fig. 1). The byproducts of the seco-acid (2) are known to be harmless (Scheme 1). Ester



Figure 1. A transition-state analog.

hydrolysis by catalytic antibodies is well established.⁶ While lactone hydrolysis has not been reported, the reverse reaction—lactone formation—has been demonstrated.⁷



Scheme 1. Hydrolysis of zearalenone (1).

Retrosynthetic analysis of the hapten, a phosphonomacrolactone,⁸ is illustrated in Scheme 2. We decided to pursue a racemic synthesis, since the geometry in the lactone region of the molecule was already significantly perturbed by the phosphonate. We felt that fidelity to the 10'S configuration

Keywords: Zearalenone; Lactone hydrolysis; Catalytic antibody.

^{*} Corresponding author at present address: Department of Chemistry, Louisiana State University, Baton Rouge, LA 70803, USA. Tel.: +1 2255784751; fax: +1 2255783458; e-mail: cmtaylor@lsu.edu

^{0040–4020/\$ -} see front matter 0 2007 Elsevier Ltd. All rights reserved. doi:10.1016/j.tet.2007.07.042

in zearalenone would serve little purpose. Indeed, the use of both enantiomers, in antibody generation, would double our chances of inducing an effective complementary binding site.



Scheme 2. Retrosynthetic analaysis.

Compound 3 represents an advanced intermediate that requires deprotection, followed by conjugation of the ketone to a carrier protein.⁹ Compound **3** is conveniently broken down into an aromatic fragment 4 and a aliphatic fragment 5. It was envisaged that formation of a phosphonate ester would precede formation of the 14-membered ring via an RCM reaction. The latter has precedent in a recent synthesis of zearalenone itself by Fürstner et al.¹⁰ We considered a number of approaches to the synthesis of the aliphatic fragment 5. A key building block was an alkyl halide typified by iodide 6. Displacement of the halide in 6, with the anion derived from dithiane 7,11 could be expected to assemble the carbon skeleton of the aliphatic fragment. The aryl phosphonate presented two significant challenges: formation of the Ar-P bond and formation of the phosphonomacrolactone. We have recently reported the trials and tribulations associated with the synthesis of 4^{12} In this paper, we describe our synthesis of 5 and the amalgamation of the two fragments.

2. Results and discussion

The synthesis of diethyl phosphonate **11** was reported elsewhere¹² and is summarized in Scheme 3. The only low-yielding step was the conversion of C3-symmetric phloroglucinol (**8**) to its diisopropyl ether derivative. Phosphorylation of the remaining phenol was conducted under Atherton–Todd conditions.¹³ An anionic phospho-Fries rearrangement¹⁴ led to compound **10**. The phenol in compound **10** was converted to the corresponding triflate and then to the styrene **11**, under Stille conditions. Partial hydrolysis of the phosphonate ester gave compound **4**.



Scheme 3. Synthesis of the aromatic fragment.

We prepared iodide **6**, and the corresponding bromide **15**, in racemic form, according to Scheme 4. Hydrolysis of the lactone in γ -valerolactone (**12**) and protection of the sodium salt gave the bis-TBDMS derivative **13**. The ester was reduced to the primary alcohol **14** by analogy to a procedure reported by Amino et al.¹⁵ The bromide **15** had been prepared and utilized previously,¹⁶ but in our hands it was unstable and difficult to purify. Fortunately, iodide **6** was readily obtained; although it was best to prepare this compound immediately prior to coupling.



Scheme 4.

Dithiane 7 has been prepared previously by reaction of the anion of 1,3-dithiane (16) with 5-bromopent-1-ene (Scheme 5).¹⁷ Careful optimization of reaction conditions led to the formation of 7 in ~70% yield, but it was incredibly difficult to separate 7 from residual 16 and byproducts of the butyllithium. An alternative synthesis, via hex-5-enal,¹⁸ gave a lower yield of dithiane 7, but it could be produced on gram scale in high purity. Condensation of the anion of 7 with iodide 6 gave compound 18^{19} in excellent yield. The secondary alcohol was liberated under standard conditions to give 5, ready for coupling to the aromatic fragment.

Prior to committing our valuable intermediates **4** and **5**, we investigated the assembly of a phosphonomacrolactone in a model system (Scheme 6). There are many methods for phosphonate ester formation, but recent reports encouraged us to use Mitsunobu conditions.^{8b} Thus, condensation of the simplified phosphonate 19^{12} and dec-9-en-1-ol via a Mitsunobu reaction²⁰ gave **20** in moderate yield. Closure of the



Scheme 5. Assembly of the aliphatic fragment.



Scheme 6. Model system for macrocycle formation.

14-membered ring to give **21** was accomplished in excellent yield via standard RCM conditions.

Turning our attention to the more complex system, we encountered considerable difficulties (Scheme 7). The best yield obtained for the Mitsunobu reaction was 26% and this required the use of 2 equiv of the acid (so-called alcohol-limiting conditions). Previous experience²¹ encouraged us to prepare the acid chloride of **4**, but this failed to condense at all with alcohol **5**, even in the presence of silver cyanide.

The RCM step was also much lower yielding than in the model system. We suspected that the dithiane functionality may be the cause of problems in one or both of these crucial steps. We therefore removed the dithiane from alcohol **5** (HCl, DMSO) and performed the esterification and RCM reactions with no improvements in yields.



Scheme 7. Formation of the hapten macrocycle.

3. Conclusion

We have prepared compound 23, an advanced intermediate *en route* to a transition-state analog for the hydrolysis of the lactone functionality in zearalenone. The phosphonic acid was introduced onto the aromatic ring via an anionic phospho-Fries rearrangement. Phosphonate ester formation, via a Mitsunobu reaction, gave a disappointing yield in the formation of model system 20 and this deteriorated in the real system 22. Ring closing metathesis was an effective means of forming the macrocycle in zearalenone itself¹⁰ and the model system 21 (Scheme 6). The yield was low in the real system. Nevertheless, sufficient material can be produced in a convergent manner, to investigate the generation of antibodies that might catalyze the degradation of zearalenone.

4. Experimental

4.1. General details

All reactions were conducted under a dry nitrogen atmosphere unless otherwise noted. Reagents were obtained from commercial suppliers and used directly with the following exceptions. Tetrahydrofuran was dried over sodium/benzophenone and distilled prior to use. Diisopropylethylamine, triethylamine, and pyridine were dried and distilled from CaH₂ and stored over KOH pellets. Flash chromatography was performed using Scharlau 60 silica gel (230–400 mesh) with the indicated solvents. Thin layer chromatography (TLC) was carried out on precoated silica plates (Merck Kieselgel 60 F254) and compounds were visualized by UV fluorescence or by staining with anisaldehyde or phosphomolybdic acid. ¹H and ¹³C NMR spectra were obtained using either a JEOL JNM-GX270W or a Bruker Avance 400 spectrometer. Chemical shifts for spectra in CDCl₃ are given in parts per million (ppm) downfield from tetramethylsilane as internal standard (¹H) or relative to residual solvent (13C). High resolution mass spectra were recorded using a VG7070 mass spectrometer operating at nominal accelerating voltage of 70 eV.

4.1.1. (2,4-Diisopropoxy-6-vinyl-phenyl)phosphonic acid monoethyl ester (4). A solution of diester derivative 11 (100 mg, 0.8 mmol, 1 equiv) in EtOH (2 mL) and 2 M NaOH (2 mL) was heated at reflux for 3 h. The mixture was concentrated to remove the ethanol, diluted with water (8 mL), neutralized with concd HCl, and extracted with EtOAc (2×15 mL). The combined organic extracts were washed with water (20 mL), dried over MgSO₄, filtered, and concentrated to give 4 as a colorless oil (191 mg, 90%). This crude product was used without further purification. R_f 0.25 (10:1 CH₂Cl₂-MeOH). ¹H NMR (CDCl₃, 400 MHz) δ 1.28 (3H, t, J 7.0 Hz, OCH₂CH₃), 1.35 (6H, d, J 6.0 Hz, CHMe₂), 1.39 (6H, d, J 6.0 Hz, CHMe₂), 4.04 (2H, app.p, J 7.0 Hz, POCH₂CH₃), 4.62 (2H, hept., J 6.0 Hz, 2×CHMe₂×2), 5.29 (1H, d, J 10.8 Hz, CH=CH₂ cis), 5.51 (1H, dd, J 17.2, 1.4 Hz, CH=CH₂ trans), 6.39 (1H, dd, J 5.5, 2.3 Hz, ArH), 6.66 (1H, dd, J 5.5, 2.3 Hz, ArH), 7.68 (1H, dd, J 17.2, 10.8 Hz, CH=CH₂); ¹³C NMR (CDCl₃, 100 MHz) δ 16.2 (d, J 7 Hz), 21.8, 22.0, 61.2 (d, J 6 Hz), 69.9, 71.6, 101.2 (d, J 10 Hz), 106.6 (d, J 15 Hz), 107.7 (d, J 193 Hz), 116.0, 137.7 (d, J 4 Hz), 146.2 (d, J 10 Hz), 161.7, 161.7 (d, J 6 Hz); HRMS (EI⁺): M⁺, found 328.14442. C₁₆H₂₅O₅P requires 328.14396.

4.1.2. (±)-4-tert-Butyldimethylsilyloxypentanoic acid tertbutyldimethylsilyl ester (13). γ -Valerolactone (12) (2.60 g, 26 mmol, 1.0 equiv) was dissolved in a mixture of water (15 mL) and dioxane (15 mL). Sodium hydroxide (1.038 g, 26 mmol, 1.0 equiv) was added and the solution stirred at rt for 30 min. The mixture was concentrated and dried over P_2O_5 to give the sodium salt of 4-hydroxypentanoic acid as a colorless solid. This was suspended in DMF (60 mL). tert-Butyldimethylsilylchloride (11.75 g, 78 mmol, 3.0 equiv), imidazole (7.08 g, 104 mmol, 4.0 equiv), and DMAP (1.59 g, 13 mmol, 0.50 equiv) were added. The mixture was stirred at rt overnight and then at 50 °C for 1 h. Water (60 mL) was added and the mixture extracted with diethyl ether $(3 \times 35 \text{ mL})$. The combined organic layers were washed with 10% citric acid (20 mL) and brine (20 mL), dried over MgSO₄, filtered, and concentrated. The residue was purified by distillation to give 13 as a colorless oil (6.84 g, 91%). R_f 0.72 (3:1 hexanes-EtOAc); bp 110 °C (0.1 mmHg). ¹H NMR (CDCl₃, 270 MHz) δ 0.04 (6H, s, SiMe₂), 0.26 (6H, s, SiMe₂), 0.88 (9H, s, Si^tBu), 0.93 (9H, s, Si^tBu), 1.13 (3H, d, J 6.2 Hz, MeCHOTBDMS), 1.68 (2H, m, CH2CH2COOTBDMS), 2.38 (2H, t, J 7.5 Hz, CH₂COOTBDMS), 3.86 (1H, m, CHOTBDMS); ¹³C NMR (CDCl₃, 67.5 MHz) & -4.8, -4.4, 17.6, 18.1, 23.8, 25.6, 25.9, 32.0, 34.6, 67.4, 174.1; HRMS (FAB+): MH+, found 347.2428451. C₁₇H₃₉O₃Si₂ requires 347.243777.

4.1.3. (±)-4-*tert*-Butyldimethylsilyloxy-pentan-1-ol (14). A solution of **13** (910 mg, 2.6 mmol, 1.0 equiv) in diethyl ether (3 mL) was added dropwise over 10 min to a suspension of LiAlH₄ (220 mg, 5.8 mmol, 2.2 equiv) in diethyl ether (7.4 mL) at 0 °C. The mixture was stirred at 0 °C for 1 h 45 min and then at rt for 10 min. The reaction was quenched by the addition of a few drops of methanol. The mixture was diluted with EtOAc (70 mL) and washed with brine (35 mL). The combined aqueous layers were extracted with EtOAc (20 mL). The combined organic extracts were dried over MgSO₄, filtered, and concentrated. The residue was purified by flash chromatography, eluting with 10:1

hexanes–EtOAc to give **14** (620 mg, 52%). R_f 0.33 (3:1 hexanes–EtOAc). ¹H NMR (CDCl₃, 270 MHz) δ 0.07 (6H, s, Si Me_2), 0.89 (9H, s, Si'Bu), 1.16 (3H, d, J 6.2 Hz, Me-CHOTBDMS), 1.5–1.7 (4H, m, CH₂CH₂CH₂OH), 3.63 (2H, m, CH₂OH), 3.89 (1H, m, CHOTBDMS); ¹³C NMR (CDCl₃, 100 MHz) δ –4.8, –4.5, 18.1, 23.2, 25.8, 28.5, 36.0, 63.0, 68.4; HRMS (FAB⁺): MH⁺, found 219.1777. C₁₁H₂₇O₂Si requires 219.1780.

4.1.4. (±)-2-tert-Butyldimethylsilyloxy-5-iodopentane (6). Triphenvlphosphine (528 mg, 2 mmol, 2 equiv), imidazole (204 mg, 3 mmol, 3 equiv), and iodine (764 mg, 3 mmol, 3 equiv) were added sequentially to a solution of primary alcohol 14 (218 mg, 1 mmol, 1 equiv) in THF (10 mL) at 0 °C under nitrogen. The mixture was warmed to rt and stirred for 3 h. The mixture was filtered through a pad of silica gel and the filtrate was concentrated. The residue was purified by flash chromatography, eluting with 10:1 hexanes-EtOAc to give the iodide derivative 6 as a colorless oil (308 mg, 94%). R_f 0.60 (10:1 Hex-EtOAc). ¹H NMR (CDCl₃, 400 MHz) δ 0.03 (6H, s, SiMe₂), 0.87 (9H, s, Si^tBu), 1.11 (3H, d, J 6.0 Hz, MeCHOTBDMS), 1.46-1.50 (2H, m, CH₂CH₂CH₂I), 1.79–1.93 (2H, m, CH₂CH₂I), 3.17 (2H, t, J 7.0 Hz, CH₂I), 3.80 (1H, sext., J 6.0 Hz, CHOTBDMS); ¹³C NMR (CDCl₃, 100 MHz) δ -4.8, -4.4, 7.3, 18.0, 23.8, 25.8, 29.8, 40.3, 67.6; HRMS (EI+): MH⁺, found 329.07945. C₁₁H₂₆IOSi requires 329.07977.

4.1.5. Hex-5-enal.²² Molecular sieves (2.5 g, 4 Å) were added to a solution of 5-hexen-1-ol (1.00 g, 10 mmol, 1.0 equiv) in CH₂Cl₂ (100 mL) at 0 °C. The mixture was stirred for 5 min. Pyridinium chlorochromate (3.23 g. 15 mmol, 1.5 equiv) was added in portions over a period of 5 min, and the mixture was stirred at rt for 4 h. The reaction mixture was diluted with diethyl ether (100 mL) and filtered through a mixture of silica gel, Celite[™], and activated charcoal. The dark brown residue was washed with diethyl ether (40 mL). The greenish yellow solution was concentrated to ~ 10 mL and transferred to a round bottomed flask. Short-path distillation led to the isolation of hex-5-enal as a colorless liquid (325 mg, 32%). Bp 38 °C (1 atm). ¹H NMR (CDCl₃, 400 MHz) δ 1.72 (2H, pent., J 7.3 Hz, CH₂CH₂CH₂), 2.07 (2H, q, J 7.3 Hz, CH₂CH=CH₂), 2.45 (2H, td, J 7.3, 1.6 Hz, CH₂CH=O), 4.90-5.09 (2H, m, CH=CH₂), 5.74 (1H, ddt, J 17.0, 10.7, 7.3 Hz, $CH=CH_2$), 9.74 (1H, d, J 1.6 Hz, CH=O); ¹³C NMR (CDCl₃, 100 MHz) δ 21.1, 32.9, 43.0, 115.5, 137.5, 202.5.

4.1.6. 2-(4-Pentenyl)-1,3-dithiane (7). Propane-1,3-dithiol (101 μ L, 108 mg, 1 mmol, 1 equiv) was added to a solution of hex-5-en-al (100 mg, 1 mmol, 1 equiv) in CHCl₃ (2 mL) at rt, stirred for 1 h, and then cooled to $-10 \,^{\circ}$ C. BF₃·OEt₂ (128 μ L, 144 mg, 1 mmol, 1 equiv) was added dropwise and the resulting solution was allowed to warm to rt and stirred for 17 h. The mixture was washed with water (3×5 mL), 10% aqueous NaOH (5 mL), and water (5 mL), dried over MgSO₄, filtered, and concentrated. The residue was purified by flash chromatography, eluting with 10:1 hexanes–EtOAc to give the dithiane derivative 7 as a colorless oil (177 mg, 94%). R_f 0.43 (10:1 Hex–EtOAc). ¹H NMR (CDCl₃, 400 MHz) δ 1.55–1.63 (2H, m, CH₂CH=CH₂), 1.71–1.79 (2H, m, CH₂CH=CH₂), 1.80–1.88 (2H, m, CH₂CHS), 2.03–2.12 (2H, m, CH₂CH=CH₂), 2.77–2.90 (4H, m,

10081

CH₂S×2), 4.02 (1H, t, J 7.0 Hz, SCHS), 4.93–5.03 (2H, m, CH=CH₂), 5.72–5.82 (1H, m, CH=CH₂); ¹³C NMR (CDCl₃, 100 MHz) δ 25.7, 26.0, 30.4 (2C), 33.2, 34.8, 47.4, 114.9, 138.0; HRMS (EI): M⁺, found 188.06936. C₉H₁₆S₂ requires 188.06934.

4.1.7. 2-tert-Butyldimethylsilyloxy-10-undecen-6-1,3-dithiane (18).¹⁹ "BuLi (350 µL of 1.5 M solution in hexane, 33.6 mg, 0.525 mmol, 1.05 equiv) was added dropwise to a stirred solution of dithiane 7 (94 mg, 0.5 mmol, 1.00 equiv) in THF (3 mL) at -20 °C under nitrogen. The mixture was stirred at -20 °C for 2 h. The temperature was decreased to -78 °C and a solution of freshly prepared iodide 6 (180 mg, 0.55 mmol, 1.10 equiv) in THF (2 mL) was added dropwise. The temperature was gradually warmed to -20 °C and stirred for 3 h. The reaction was quenched with aq NaHCO₃ (2 mL) and extracted with diethyl ether (2×15 mL). The combined organic layers were washed with water (15 mL) and brine (15 mL), dried over MgSO₄, filtered, and concentrated. The residue was purified by flash chromatography, eluting with 20:1 hexanes-EtOAc to give dithiane derivative 18 as a colorless oil (181 mg, 93%). R_f 0.48 (10:1 Hex-EtOAc). ¹H NMR (CDCl₃, 400 MHz) δ 0.06 (3H, s, SiMe), 0.07 (3H, s, SiMe), 0.90 (9H, s, Si^tBu), 1.14 (3H, d, J 6.1 Hz, MeCHOTBDMS), 1.22-1.52 (6H, m), 1.83-1.99 (6H, m), 2.08 (2H, q, J 7.0 Hz, CH₂CH=CH₂), 2.79–2.82 (4H, m, CH₂S×2), 3.81 (1H, app. sext., J 6.1 Hz, CHOTBDMS), 4.97-5.07 (2H, m, $CH = CH_2$), 5.80 (1H, ddt, J 17.2, 10.2, 7.0 Hz, CH=CH₂); ¹³C NMR (CDCl₃, 100 MHz) δ -4.7, -4.4, 18.1, 20.4, 23.4, 23.8, 25.6, 25.9, 33.7, 37.4, 38.4, 39.8, 53.2, 68.5, 115.0, 138.3; HRMS (EI): M⁺, found 388.23008. C₂₀H₄₀OS₂Si requires 388.22899.

4.1.8. 2-(4-Hydroxypentyl)-2-(4-pentenyl)-1,3-dithiane (5). TBAF (220 µL, 1 M solution in THF, 0.22 mmol, 1.1 equiv) was added to a solution of silyl ether 18 (78 mg, 0.2 mmol, 1.0 equiv) in THF (1 mL) at rt under nitrogen. The mixture was stirred overnight, and then filtered through a pad of silica gel. The filtrate was concentrated and the residue purified by flash column chromatography, eluting with 5:1 hexanes-EtOAc to give alcohol 5 as a colorless oil (40 mg, 72%). R_f 0.13 (5:1 Hex-EtOAc). ¹H NMR (CDCl₃, 400 MHz) δ 1.16 (3H, d, J 6.2 Hz, MeCHOH), 1.39-1.53 (6H, m), 1.83-1.92 (6H, m), 2.06 (2H, q, J 7.0 Hz, CH₂CH=CH₂), 2.77-2.78 (4H, m, CH₂S×2), 3.78 (1H, sext., J 6.2 Hz, CHOH), 4.96 (1H, dd, J 10.2, 2.0 Hz, CH=CH₂ cis), 5.00 (1H, dd, J 17.2, 2.0 Hz, CH=CH₂ trans), 5.78 (1H, ddt, J 17.2, 10.2, 7.0 Hz, CH=CH₂); ¹³C NMR (CDCl₃, 100 MHz) δ 20.4, 23.3, 23.5, 25.4, 25.9 (2C), 33.6, 37.5, 38.1, 39.2, 53.1, 67.8, 115.0, 138.2; HRMS (EI): M^+ , found 274.14275. $C_{14}H_{26}OS_2$ requires 274.14251.

4.1.9. Ethyl non-8-enyl (2-vinylphenyl)phosphonate (20). A solution of dec-9-en-1-ol (156 μ L, 137 mg, 0.88 mmol, 1.1 equiv) and triphenylphosphine (231 mg, 0.88 mmol, 1.1 equiv) in THF (8 mL) was added dropwise to a solution of acid **19** (170 mg, 0.8 mmol, 1.0 equiv) and dimethylazo-dicarboxylate (174 μ L, 128 mg, 0.88 mmol, 1.1 equiv) in THF (8 mL) at rt. The mixture was stirred under nitrogen for 4 h, concentrated, and the residue purified by flash chromatography, eluting with hexanes–EtOAc, with solvent

gradient from 10:1 to 5:1 to give the ester derivative 20 as a colorless oil (135 mg, 48%). Rf 0.25 (2:1 Hex-EtOAc). ¹H NMR (CDCl₃, 400 MHz) δ 1.18–1.36 (13H, m, CH₂×5 and OCH₂CH₃), 1.58-1.66 (2H, m, POCH₂CH₂), 1.96-2.06 (2H, m, CH₂CH=CH₂), 3.92-4.16 (4H, m, POCH₂×2), 4.86–5.02 (2H, m, CH₂CH=CH₂), 5.35 (1H, d, J 10.8 Hz, ArCH=CH₂ cis), 5.70 (1H, d, J 17.4 Hz, ArCH= CH_2 trans), 5.74–5.82 (1H, m, CH₂CH= CH_2), 7.29-7.44 (2H, m, ArH), 7.49 (1H, t, J 7.0 Hz, ArH), 7.63 (1H, t, J 7.0 Hz, ArH), 7.3 (1H, dd, J 17.4, 10.8 Hz, ArCH=CH₂): ¹³C NMR (CDCl₃, 100 MHz) δ 16.2 (d, J 7 Hz), 25.4, 28.8, 28.9, 29.0, 29.2, 30.3 (d, J 7 Hz), 33.7, 62.0 (d, J 5 Hz), 66.0 (d, J 5 Hz), 114.0, 116.7, 125.6 (d, J 181 Hz), 126.1 (d, J 15 Hz), 127.1 (d, J 15 Hz), 134.0 (d, J 3 Hz), 134.1 (d, J 10 Hz), 135.3 (d, J 5 Hz), 139.1, 141.2 (d, J 10 Hz); HRMS (EI): M⁺, found 350.20067. C₂₀H₃₁O₃P requires 350.20108.

4.1.10. 2-Ethoxy-(2,3-benzo-1-oxa-2-phosphacyclotetradec-5-ene) 2-oxide (21). Grubbs' second generation catalyst (12 mg, 0.014 mmol, 5 mol %) was added to a solution of ester 20 (100 mg, 0.28 mmol, 1 equiv) in toluene (70 mL). The mixture was stirred at 80 °C under nitrogen for 23 h, concentrated, and the resulting residue purified by flash chromatography, eluting with 2:1 hexanes-EtOAc to give 21 as a colorless oil (82 mg, 90%). $R_f 0.13$ (2:1 Hex–EtOAc). ¹H NMR (CDCl₃, 400 MHz) δ 1.23–1.63 (12H, m, CH₂×6), 1.25 (3H, t, J 6.8 Hz, POCH₂CH₃), 2.24 (2H, q, J 6.5 Hz, CH=CHC H_2), 3.91–4.16 (4H, m, POC $H \times 2$), 6.12 (1H, dt, J 15.7, 6.5 Hz, ArCH=CH), 7.03 (1H, d, J 15.7 Hz, ArCH=CH), 7.18-7.23 (1H, m, ArH), 7.40 (1H, t, J 7.3 Hz, ArH), 7.56 (1H, t, J 7.3 Hz, ArH), 7.91 (1H, dd, J 14.8, 7.3 Hz, ArH); ¹³C NMR (CDCl₃, 100 MHz) δ 16.3 (d, J 6 Hz), 22.9, 24.5, 25.7, 26.1, 26.2, 29.1 (d, J 7 Hz), 31.0, 62.3 (d, J 5 Hz), 64.9 (d, J 7 Hz), 124.8 (d, J 185 Hz), 125.8 (d, J 14 Hz), 126.1 (d, J 14 Hz), 128.4 (d, J 5 Hz), 132.5 (d, J 3 Hz), 133.7, 134.2 (d, J 10 Hz), 141.3 (d, J 9 Hz); HRMS (EI): M⁺, found 322.16973. C₁₈H₂₇O₃P requires 322.16978.

4.1.11. Ethyl 2-[6-(1,3-dithiane)undec-10-enyl] (4,6-diisopropoxy-2-vinylphenyl)phosphonate (22). A solution of alcohol 5 (60 mg, 0.22 mmol, 1.0 equiv) and triphenylphosphine (58 mg, 0.22 mmol, 1.0 equiv) in THF (2 mL) was added dropwise to a solution of acid 4 (144 mg, 0.44 mmol, 2.0 equiv) and dimethylazodicarboxylate (32 mg, 0.22 mmol, 1.0 equiv) in THF (3 mL) at rt. The mixture was stirred under an atmosphere of nitrogen for 3 h, concentrated, and the residue purified by flash chromatography, eluting with hexanes-EtOAc with a gradient from 10:1 to 5:1 to give the ester derivative 22 as a colorless oil (33 mg, 26%). R_f 0.28 (2:1 Hex–EtOAc). ¹H NMR (CDCl₃, 400 MHz) δ 1.16 (3H, d, J 6.2 Hz, CH(OR)Me), 1.25 (3H, td, J 7.0, 4.4 Hz, POCH₂CH₃), 1.31 (12H, d, J 6.0 Hz, OCHMe₂×2), 1.37-2.06 (14H, m), 2.65-2.77 (4H, m, CH₂S×2), 3.90-4.13 (2H, m, POCH₂), 4.48-4.63 (3H, m, CHMe₂×2 and CH(OR)Me), 4.90-5.05 (2H, m, CH₂CH=CH₂), 5.23 (1H, d, J 10.8 Hz, ArCH=CH₂ cis), 5.44 (1H, dd, J 17.2, 1.6 Hz, ArCH=CH₂ trans), 5.73 (1H, ddt, J 17.2, 10.8, 6.8 Hz, CH₂CH=CH₂), 6.31 (1H, d, J 4.0 Hz, ArH), 6.52 (1H, dd, J 4.0, 2.4 Hz, ArH), 7.81 (1H, dd, J 17.2, 10.8 Hz, ArCH=CH₂); ¹³C NMR (CDCl₃, 100 MHz) δ 16.2, 20.0, 21.5, 22.0 (2C), 22.1 (3C), 23.3, 25.6, 26.0, 33.8, 37.7,

38.2, 53.0, 60.9, 61.0, 69.7, 70.4, 73.0, 100.5, 105.7, 107.2 (*J* 154 Hz), 114.9, 115.5, 138.1, 138.2, 147.0, 161.6, 162.0; HRMS (EI): M⁺, found 584.27636. C₃₀H₄₉O₅PS₂ requires 584.27591.

4.1.12. 2-Ethoxy-2,3-(1',5',-diisopropoxy)benzo-10-(1,3dithiane)-14-methyl-1-oxa-2-phosphacyclotetradec-5ene 2-oxide (23). Grubbs' second generation catalyst (8.5 mg, 0.01 mmol, 10 mol %) was added to a solution of ester 22 (58.5 mg, 0.1 mmol, 1 equiv) in toluene (50 mL). The reaction mixture was stirred at 80 °C under nitrogen for 23 h, concentrated, and the resulting residue purified by flash chromatography, eluting with 2:1 hexanes-EtOAc to give macrolactone 23 as a colorless oil (27 mg, 48%). R_f 0.30 (2:1 Hex–EtOAc). ¹H NMR (CDCl₃, 400 MHz) δ 1.28–1.36 (18H, m, MeCHO, POCH₂CH₃, OCHMe₂×2), 1.47-2.37 (14H, m), 2.73-2.83 (4H, m, CH₂S×2), 3.84-3.99 (1H, POCH₂), 4.02–4.15 (1H, m, POCH₂), 4.50 (1H, pent. J 6.0 Hz, OCHMe₂), 4.58 (1H, pent, J 6.0 Hz, OCHMe₂), 4.72–4.81 (1H, m, MeCHO), 5.94 (1H, dt, J 15.8, 7.1 Hz, ArCH=CH), 6.28 (1H, dd, J 5.0, 2.2 Hz, ArH), 6.49 (1H, dd, J 5.0, 2.2 Hz, ArH), 7.56 (1H d, J 15.8 Hz, ArCH=CH); ¹³C NMR (CDCl₃, 100 MHz) δ 12.7, 16.4, 21.9 (2C), 22.0 (4C), 23.9, 25.7, 26.0, 30.2, 35.6, 37.2, 37.5, 53.4, 61.2, 61.3, 69.7, 70.8, 71.3, 100.6, 107.3 (d, J 193 Hz), 115.1, 132.5, 147.5, 161.5, 161.6; HRMS (EI): M⁺, found 556.24517. C₂₈H₄₅O₅PS₂ requires 556.24461.

Acknowledgements

We thank the Massey University Research Fund (MURF) for a postdoctoral fellowship to C.M.T and K.P.J. We thank Drs. Chris Miles, Ian Garthwaite, Mark Dines, and Neale Towers of AgResearch (Ruakura, NZ) for helpful discussions.

Supplementary data

¹H and ¹³C NMR spectra for compounds **4**, **5**, **6**, **7**, **13**, **14**, **18**, **20**, **21**, **22**, and **23**. Supplementary data associated with this article can be found in the online version, at doi:10.1016/j.tet.2007.07.042.

References and notes

- (a) Stob, M.; Baldwin, R. S.; Tuite, J.; Andrews, F. N.; Gillette, K. G. *Nature* **1962**, *196*, 1318; (b) Urry, W. H.; Wehrmeister, H. L.; Hodge, E. B.; Hidy, P. H. *Tetrahedron Lett.* **1966**, *27*, 3109–3114.
- (a) Placinta, C. M.; D'Mello, J. P. F.; Macdonald, A. M. C. Animal Feed Sci. Technol. 1999, 78, 21–37; (b) Newsome, R. Food Technol. 2006, 60, 51–58; (c) Bennett, J. W.; Klich, M. Clin. Microbiol. Rev. 2003, 16, 497–516; (d) Conkova, E.; Laciaková, A.; Kovac, G.; Seidel, H. Vet. J. 2003, 165, 214–220.
- (a) De Menna, M. E.; Lauren, D. R.; Poole, P. R.; Mortimer, P. H.; Hill, R. A.; Agnew, M. P. NZ J. Agr. Res. 1987, 30, 499–504; (b) Towers, N. R. 19th German Mycotoxin

Workshop, Munich, June 2–4, 1997; (c) Towers, N. R.; Sprosen, J. M. *NZ Vet. J.* **1993**, *41*, 223–224.

- 4. (a) El-Sharkaway, S.; Abdul-Hajj, Y. J. *Xenobiotica* 1988, *18*, 365–371; (b) Megharaj, M.; Garthwaite, I.; Thiele, J. H. *Lett. Appl. Microbiol.* 1997, *24*, 329–333.
- http://www.agvax.com/animal_health/sheep/androvax/index. htm.
- For example, antibodies have been developed for the hydrolysis of cocaine: Landry, D. W.; Zhao, K.; Yang, G. X.-Q.; Glickman, M.; Georgiadis, T. M. *Science* **1993**, *259*, 1899–1901.
- (a) Formation of a six-membered ring: Napper, A. D.; Benkovic, S. J.; Tramontano, A.; Lerner, R. A. *Science* 1987, 237, 1041–1043; (b) Formation of a 14-membered ring: Pungente, M. D.; Weiler, L.; Ziltener. *Can. J. Chem.* 2002, 30, 1643–1645.
- For previous syntheses of phosphonomacrolactones, see: (a) Smith, W. W.; Bartlett, P. A. J. Am. Chem. Soc. 1998, 120, 4622–4628; (b) Pungente, M. D.; Weiler, L. Org. Lett. 2001, 3, 643–646.
- Zearalenone has been conjugated to a carrier protein via the ketone previously, generating antibodies which are used in an ELISA assay for the toxin: Dixon, D. E.; Warner, R. L.; Ram, B. P.; Hart, L. P.; Pestka, J. J. J. Agric. Food Chem. 1987, 35, 122–126.
- Fürstner, A.; Thiel, O. R.; Kindler, N.; Bartkowska, B. J. Org. Chem. 2000, 65, 7990–7995.
- For a review of dithiane couplings: Smith, A. B., III; Adams, C. M. Acc. Chem. Res. 2004, 37, 365–377.
- Jayasundera, K. P.; Watson, A. J.; Taylor, C. M. Tetrahedron Lett. 2005, 46, 4311–4313.
- (a) Atherton, F. R.; Todd, A. R. J. Chem. Soc. 1947, 674–676;
 (b) Silverberg, L. J.; Dillon, J. L.; Vemishetti, P. Tetrahedron Lett. 1996, 37, 771–774.
- Taylor, C. M.; Watson, A. J. Curr. Org. Chem. 2004, 8, 623– 636.
- 15. Amino, Y.; Eto, H.; Eguchi, C. Chem. Pharm. Bull. 1989, 37, 1481–1487.
- (a) Kalivretenos, A.; Stille, J. K.; Hegedus, L. S. J. Org. Chem.
 1991, 56, 2883–2894; (b) Demeke, D.; Forsyth, C. J. Tetrahedron 2002, 58, 6531–6544.
- (a) Chung, S. K.; Dunn, L. B., Jr. J. Org. Chem. **1984**, 49, 935– 939; (b) Grig, R.; Markandu, J.; Surendrakuma, S.; Thornton-Pett, M.; Warnock, W. J. Tetrahedron **1992**, 48, 10399–10422; (c) Suenaga, K.; Araki, K.; Sengoku, T.; Uemura, D. Org. Lett. **2001**, *3*, 527–529; (d) Araki, K.; Seuenaga, K.; Sesngoku, T.; Uemura, D. Tetrahedron **2002**, 58, 1983–1995.
- It is implied that material of superior quality was obtained this way: Yamaguchi, Y.; Yamada, H.; Hayakawa, K.; Kanematsu, K. J. Org. Chem. 1987, 52, 2040–2046.
- 19. Compound 18 has been prepared via a different approach: Bracher, F.; Krauss, J. *Monatsh. Chem.* 2001, *132*, 805–811.
- For the introduction of the Mitsunobu reaction to phosphonate ester formation, see: (a) Campbell, D. A. J. Org. Chem. 1992, 57, 6331–6335; (b) Campbell, D. A.; Bermak, J. C. J. Org. Chem. 1994, 59, 658–660.
- (a) Smith, A. B., III; Taylor, C. M.; Benkovic, S. J.; Hirschmann, R. *Tetrahedron Lett.* **1994**, *35*, 6853–6856; (b) Hirschmann, R.; Yager, K. M.; Taylor, C. M.; Witherington, J.; Sprengeler, P. A.; Phillips, B. W.; Moore, W.; Smith, A. B., III. *J. Am. Chem. Soc.* **1997**, *119*, 8177–8190.
- 22. Spino, C.; Barriault, N. J. Org. Chem. 1999, 64, 5292-5298.